Pressure-dependent adhesion between solid-supported PC-lipid bilayers and vesicles under electric fields[†]

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[†] Dedicated to Professor Stanislaw (Stan) Penczek on the occasion of his 90th birthday

Abstract

Fusion of lipid bilayers in membranes is important in processes from vesicle-cell interactions (as in drug delivery) to exosome-cell signaling, while transient transmembrane electric fields are known to occur spontaneously. Two contacting phosphatidylcholine (PC) lipid membranes are known to fuse into one under external electric fields, suggesting that the interaction between them is modified by the field as they approach, prior to the fusion event. Here we measure directly the adhesion energy between dimyristoylphosphatidylcholine (DMPC) and between distearoylphosphatidylcholine (DSPC) surface layers attached to solid substrates both without and with a transmembrane electric field. We find a marked pressure-dependent adhesion behavior in the electric field, which we attribute to fusion intermediates that are formed, shedding new light on membrane electro-fusion.

Keywords: lipid bilayers; electrofusion; adhesion energy; phosphatidylcholine (PC) lipids

Introduction

Phosphatidylcholine (PC) lipids are one of the main components of the biological membrane systems, such as cell membranes, organelle membranes and exosomes, and play important roles in cell proliferative growth and programmed death [1, 2]. Liposomes or lipid nanoparticles, using PC lipid as carriers or protecting membranes, are widely used in drug delivery systems[3, 4], especially in vaccines [5]. PC-lipids are also found in healthy synovial fluids in the major joints (such as hips and knees) and on the surface of the articular cartilage layers coating the ends of joints, where they are the major lipid component (up to ca. 42% of total lipids [6, 7]). Indeed, the excellent lubrication of healthy articular cartilage, essential for the well-being of joints, has been attributed to PC lipid bilayers exposed at its outer surface [8]. In model surface forces balance (SFB) experiments, a friction coefficient down to $\mu \approx 10^{-4}$ is reported by Sorkin [9] and Goldberg [10] between PC-covered mica surfaces in physiological conditions. Such low friction is attributed, via the hydration lubrication mechanism [9], to the tenaciously-held hydration shells surrounding the PC-headgroups [11].

Tribological studies to date between lipid layers using the SFB [9, 10] were generally carried out under "zero" transverse electric fields, as the two opposing mica substrates bear identical surface charge density [12]. The presence of electric fields may considerably modify the bilayer structure (e.g. via electroporation or membrane fusion [13]) and thus the interaction between the bilayers. Moreover, recent work [14] suggests that significant transverse fields across cell membranes in vivo may arise spontaneously due to transient potential imbalance across the lipid bilayers. Any structural changes in the lipid membranes induced by such fields are expected also to influence the lateral frictional forces. Indeed in very recent studies, we found that transverse electric fields had a strong effect on the sliding friction between lipid bilayers, and this is attributed to field-induced topological changes such as electroporation [15-17].

It is also reasonable to expect that the adhesion energy, i.e. the energy required to separate two contacting lipid-bilayer bearing surfaces, might change in the presence of a transverse electric field, due to such changes in the bilayer structures, and that this would also depend on the applied pressure as this would further modulate these structures. In the present study, we explore the effects of electric field and normal pressure on adhesion between mica and gold substrates bearing dimyristoylphosphatidylcholine (DMPC) or distearoylphosphatidylcholine (DSPC) lipid layers. The DMPC lipids, close to their liquid-disordered phase at room temperature, are in the form of extended lipid bilayers (LBs) on the substrates (as their liposomes rupture when adsorbed on the surfaces), while the gel-phase DSPC vesicles retain their liposomic structure on the surfaces [9].

2. Materials and methods

2.1 Materials

Water used was purified (TOC<1 ppb and 18.2 MΩ•cm conductivity) using a BarnsteadTM GenPureTM system (Thermal Scientific). Ruby muscovite mica (grade I) was purchased from S & J Trading, Inc. Gold and silver pellets (99.999%) were obtained from Kurt J. Lesker Inc. DMPC and DSPC are purchased from Lipoid (Germany) and used as received. Platinum wire (99.99%) for electrodes is obtained from Advent Ltd. All solvents were analytical grade (Merck, Sigma, and BioLab). EPON 1004 resin (Shell) is used to glue the gold and mica on fused hemicylindrical silica lenses.

2.2 Liposome extrusion and adsorption

The DMPC and DSPC liposomes are prepared using a standard extrusion method[18]. The lipid powder is vortex-dispersed in pure water at 5 mM concentration, then sonicated for 20 min at 10 °C above respective phase transition temperature $T_{\rm M}$ ($T_{\rm M}$ = 24 °C for DMPC and 55 °C for DSPC [19]) forming dispersions of multilamellar vesicles (MLVs). The MLVs are downsized by passing the dispersion through filters with designated pore radius for several cycles: 0.4 µm for 8 cycles, 0.1 µm for 10 cycles, and 0.05 µm for 12 cycles, to yield dispersions of small unilamellar vesicles (SUVs, or liposomes). The dispersion temperature is maintained during the downsizing process following which dispersions are kept at 4 °C for less than 12 hours before use. Gold-coated or bare mica surfaces (substrates) are incubated overnight in 0.5 mM liposome dispersion then washed before mounting into the SFB.

2.3 Surface force balance (SFB) experiments

Mica surfaces are prepared by cleaving mica sheets into ca. 2.5 μ m thick single crystalline facets, as reported previously [20]. Then a ca. 60 nm thick reflective silver layer is evaporated on one side of the prepared facets. Molecularly smooth gold surfaces are prepared using the template-stripping method [21]. Briefly, a ca. 60 nm thick gold film is evaporated onto a cleaved single-crystal mica template (ca. 10 μ m thick) at the rate of 0.1 Å/min (Odem evaporator), and annealed at 150 °C for 2 hours. This gold-coated mica sheet is then glued onto the SFB lens and the molecularly smooth gold surface in contact with the atomically-smooth mica is finally exposed by peeling the template off.

The SFB experiments are carried out as described in detail previously [22, 23], using a threeelectrode modified SFB (Fig. 1). Briefly, the gold-exposing and the bare, back-silvered mica surfaces are mounted opposite to each other in a cross-cylinder configuration, with the gold surface (upper lens) mounted on a sectored piezoelectric tube and the mica substrate (lower lens), immersed in a water bath, on a normal spring set. The geometry is equivalent to a sphere (radius R ca. 1 cm) interacting with a flat surface. A beam of white light is passed through the lenses, forming multiple-beam interference fringes (fringes of equal chromatic order, as in upper insert in Fig. 1) whose wavelength is measured to yield the gold-mica separation D [24, 25]. The two surfaces are brought into contact using either a step-wise or a dynamic method and the normal forces F_n between them are calculated from the bending of the normal leaf spring. Lateral motion is induced via opposing potentials on a sectored piezoelectric (PZ) tube, and the friction force F_s measured as $F_s = K_s \Delta x$ where K_s is the shear leaf spring constant and Δx is the bending as measured by the air-gap-capacitor. The gold surface potential is controlled by a three-electrode configuration, consisting of gold as working electrode (W), and two platinum wires as counter (C) and quasi-reference electrodes (R). Normal force profiles between bare substrates are measured to calibrate the surface potential of gold using Poisson-Boltzmann equation [17]. The adhesion energy W_{adh} is calculated using JKR contacting model from the "pull-off" force $F_{pulloff}$ upon separating two surfaces [26],

$$F_{\text{pulloff}} = (3/2)\pi W_{\text{adh}}R \tag{1}$$

where R is the mean radius of curvature of the two cylindrical surfaces, measured from the fringe

shapes. In principle the adhesion energy as thus defined should be independent of the contact pressure (or contact area) between the surfaces, but for convenience we will use this definition (eq (1)) also for the present study where W_{adh} (which we may also call the apparent adhesion energy) can depend on the applied pressure.



Fig. 1: Schematic of the three-electrode-modified surface force balance (SFB) showing the main parts: sectored piezo (PZ) tube, air-capacitor probe, normal and shear spring sets. The potential of the gold surface is controlled by a three-electrode configuration: the gold surface as working electrode (W), two platinum wires as quasi-reference electrode and counter electrode. Two fused silica lenses are mounted in cross-cylinder configuration (bottom left insert) that is equivalent to a sphere on a flat configuration. The lower circular inset indicates the geometry of the gold and mica substrates. Modified with permission from ref. [27]. Copyright 2021 AIP Publishing LLC.

3. Results

The approaching (increasing load) and receding (decreasing load) normal force $F_n(D)$ profiles (normalized as $F_n(D)/R$, in the Derjaguin approximation [28]) between PC-covered substrates are measured under an electric field induced by varying the potentials Ψ_{app} applied to the gold surface; this is different to the actual potential Ψ_{gold} at the surface which can be extracted from the force profiles as earlier discussed [23]. We consider two cases, as illustrated in Fig.2.



Fig. 2: Normal force $F_n(D)/R$ profiles between two mica surfaces (a), and between gold and mica surfaces (b), collected using the dynamic method [29]. These results are used to calculate the mica surface charge density and gold surface potentials through fitting the force profiles with Poisson-Boltzmann (Gouy-Chapman) model [28] (black solid lines in a and b). The boundary conditions are constant charge vs. constant charge (panel a) and constant charge vs. constant potential (panel b). The mica surface charge σ_{mica} =-5.21 mC/m² (corresponding to $\Psi_{mica,\infty}$ =-0.095 V at infinite separation *D* in c_0 =7.8×10⁻⁵ M 1:1 electrolyte solution) is extracted from panel a and used as constant in the fitting in panel b.

This shows the normal force profiles between two bare mica surfaces (Fig. 2a), from which a surface charge density on each surface can be extracted (the mica surfaces are at constant charge), corresponding to a mica surface potential $\Psi_{\text{mica},\infty} = -0.095$ V (at infinite separation *D*). The force profiles between bare mica and a bare gold surface at different applied potentials Ψ_{app} are shown in fig. 2b: when $\Psi_{\text{app}}=-0.3$ V, the potential of the gold surface is $\Psi_{\text{gold}} = -0.089$ V, very similar to that of the bare mica, implying a low electric field $E (\approx 0)$ between them [23]. We designate that the low field case, and it is similar in terms of surface charges density and potential variation across the gap to the extensively-studied symmetric case of two interacting mica surfaces. When the applied potential is made more positive, $\Psi_{\text{app}} = -0.1$ V (red data) or 0.2 V (green data), the gold potential changes sign ($\Psi_{\text{gold}} = +0.004$ V and +0.103 respectively) and a high field $E \approx 10^7 \sim 10^8$ V/m [17] results across the gold/mica gap; this is designated the high field case.

3.1 Forces between surfaces coated with DMPC-LBs

The approaching or loading (increasing load F_n) and receding or unloading (decreasing load) normal force profiles measured at the two different potentials Ψ_{app} =-0.3 V (low field), and Ψ_{app} =-0.1 V (high field) are shown in Fig. 3a and 3b respectively. The approaching force profiles in the low field case (Fig. 3a, full symbols) show a combination of long-range (D>10 nm) electrostatic interactions and short-range steric repulsion ($D \le 10$ nm), qualitatively similarly to those in pure water between bare mica surfaces at different gold potentials (Fig. 2a). The long-range repulsions, fitted well by the Gouy-Chapman model (grey solid lines in 3a) [28], are of electrostatic origin arising from the osmotic pressure of trapped counterions. In contrast, the two oppositely charged surfaces (Fig. 3b) "jump-in" to contact at the Euler instability point when $\partial F_n/\partial D > K_n$ (black arrow in Fig. 3b). Short range interactions are dominated by steric repulsions between the DMPC-LBs, consistent with previous results[9]. Such repulsion results in a "hardwall", at separation $D = D_{hw}$ =8.6±0.8 nm (Fig. 3a) and D_{hw} =8.3±0.8 nm (Fig. 3b), indicating two confined lipid bilayers at contact [30].



Fig. 3: Typical loading and unloading normal force profiles between DMPC-LB-covered substrates, plotted as $F_n(D)/R$ in the Derjaguin approximation: a) the low-field or 'repulsive' regime (Ψ_{app} =-0.3 V, $F_n/R > 0$ on approach at all separations); and b) the high-field or 'attractive' regime (Ψ_{app} =-0.1 V, $F_n/R < 0$ on approach). Full and empty symbols represent approaching and receding profiles, respectively. Different colors represent repeat measurements, in which the maximum applied pressure (same colour code as data symbols) is different. Grey solid curves in a is the best fit using a Gouy-Chapman model [28] with mica surface charge density σ_{mica} = -5.45

mC/m², gold surface potential are the same of cases in water, Ψ_{gold} = -0.089 V (a) and Ψ_{gold} = 0.004 V (b), bulk electrolyte concentration (assuming 1:1 salt) c_0 = 7.0×10⁻⁵ M, and gold-mica Hamaker constant $A_{\rm H}$ = -9×10⁻²⁰ J [22]. The insert in b shows magnification of blue data sets.

The receding force profiles (empty symbols in Fig. 3), show both a pressure and electric field-dependent behaviour. In both low and high field cases the surfaces are compressed to different maximal loads, corresponding to differing maximal contact pressures P_{max} as indicated by the colour coding. In the low field regime (Fig. 3a) at low P_{max} , the approaching and receding profiles overlap with each other (blue data in Fig. 3a) with little hysteresis. On increasing P_{max} (Fig.3a, red and green data) there is increasing hysteresis. The possible origins of this are considered in the Discussion section. For the high *E*-field case, Fig. 3b, we observe jumps into contact as the surfaces approach, as expected [23]. Here we see that as P_{max} increases the pull-off force and thus the apparent adhesion energy increases. We note, importantly, that the data in Figs. 3a, b were all taken at a given contact point for each lipid type (to maintain the constant electric field at given potential), and that profiles were carried out successively going from lower to higher P_{max} , with ca. 15' wait time between successive loading cycles. The fact that the subsequent loading profiles reproduce themselves with little hysteresis (i.e. red data following the blue, and green following the red in Fig. 3a; and likewise in Fig. 3b with additionally orange following the green) suggests that the lipid layers largely recover in between loading processes from any compression induced changes at P_{max} , as further discussed below. Such measurements are carefully repeated also at different contact points, and similar results are acquired as summarized in Fig.4.

The adhesion energy (shown in Fig. 4), calculated from the pull-off forces according to eq (1), can be separated into two regimes. For the 'repulsive' low-field case (from data such as in Fig. 3a) there is no net adhesion at lower P_{max} values (i.e. no pull-off force from adhesion; $W_{adh} = 0$ and is independent of P_{max} ; empty symbols in Fig. 4), while at highest P_{max} there is substantial scatter in the pull-off force (and W_{adh}). In contrast, for the high field case (from data such as in Fig. 3b) the adhesion energy rises with the pressure P_{max} (full symbols in Fig.4), showing pressure dependent behavior.

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Fig. 4: Apparent adhesion energies evaluated from data such as in Fig. 3 and eq (1) between DMPC bilayer-covered gold and mica surfaces, in high electric field (Ψ_{app} = -0.1 V, full symbols), and low field (Ψ_{app} = -0.3 V, empty symbols). The data from repeat experiments at different contact points are shown in different colors. The vertical dashed lines represent the pressure for hemifusion at Ψ_{app} = -0.3 V, and the cross-hatched band shows its standard deviation.

3.2 Forces between surfaces coated with DSPC liposomes

The approaching force profiles of DSPC are monotonically repulsive (Fig.5, full symbols), irrespective of the electric field, similar to the previous results between two mica substrates [31], attributed to steric repulsion between the intact liposomes that is not influenced by the electric field [32, 33]. A "hard-wall" separation $D_{hw} = 20.4\pm0.8$ nm and 20.0 ± 0.8 nm, corresponding to 2 layers of flattened liposomes, i.e. 4 layers of LBs, are measured, indicating the integrity of DSPC-liposomes [34, 35]. The DSPC-LBs thickness is given by $D_{hw}/4 = 5.1\pm0.2$ nm and 5.0 ± 0.2 nm (compared with the corresponding DMPC-LB thickness values of 4.3 ± 0.2 nm and 4.2 ± 0.2 nm respectively).

The receding force profiles of DSPC-liposomes (empty symbols in Fig. 5) show a similar hysteresis to those of DMPC-LBs. The loading/unloading hysteresis gradually appears as the maximum pressure P_{max} increases, for DSPC-liposome both with and without electric field. This trend is observed for the low field, but is most clearly seen in the increasing pull-off forces when separating two surfaces in the high field case (Fig. 5b): the pull-off forces increase from zero (in the absence of an *E*-field, Fig. 5a) to finite increasing values as the pressure increases in the loading force profiles.



Fig. 5: Typical approaching (filled symbols) and receding (empty symbols) normal force profiles between DSPC-SUVs covered gold and mica surfaces, plotted as $F_n(D)/R$ in the Derjaguin approximation where *R* is the mean curvature radius of the surfaces: a) the low-field or 'repulsive' regime (Ψ_{app} = -0.3 V); and b) the high-field or 'attractive' regime (Ψ_{app} = 0.2 V), measured at a given contact point to maintain the constant electric field at given potential. The maximum pressures at all loading profiles are represented by different colors, and its effect is summarized below.

The adhesion energies of DSPC-liposomes are plotted against the maximum pressures P_{max} , as shown in Fig. 6. Similar to the DMPC-LBs, two different adhesion types are clearly shown. At low electric field (empty symbols), the DSPC-liposome adhesion energy $W_{\text{adh}} = 0$ and is independent of P_{max} , consistent with earlier studies [10, 31]. At high electric field (full symbols), a P_{max} -dependent apparent adhesion energy is clearly seen from the data, similar to those in DMPC-LBs (Fig. 3b and full symbols in Fig. 4).



Fig. 6: Adhesion energies (evaluated from pull-off forces as in Fig. 5, via eq (1)) between DSPC-SUVs covered gold and mica surfaces, at attractive regime (Ψ_{app} = 0.2 V, full points), and repulsive regime (Ψ_{app} = -0.3 V, empty points). The data from repeat experiments at different contact points are shown in different colors. The results indicate that the adhesion energy increases with the maximum pressure when electropores appear under electric field.

4. Discussion

The main new finding in this study is the hysteretic pressure-dependent adhesion of PCcovered substrates, weakly in the case of low or zero field between the surfaces, and much more marked when a high field (ca. $10^7 - 10^8$ V/m) is applied between them [17, 36]. Pressuredependent adhesion is common between *polymer*-coated surfaces (no electric field), due to the contacting of asperities from opposite surfaces as pressure increases [37-39]. A closer analogy for the case of zero transverse *E*-field with lipid layers is proposed by Sun et al. [40] and other groups [41-43] to explain adhesion between giant unilamellar vesicles and tethered LBs. They divide the adhesive behavior into a "low adhesion" regime, where no fusion intermediates forms, thus the adhesion is pressure-independent, and a "high adhesion" regime, where two LBs are connected by fusion intermediates (though they do not specify the nature of such 'fusion intermediates'). They attribute the stronger adhesion at higher normal pressures to the more favorable formation of fusion intermediates [40], thus additional energy is needed to separate two LBs, leading to pressure-dependent adhesion. These studies were all carried out in the absence of any applied electric fields, and are in line with our results as in Figs. 3a and 5a with no electric fields across the bilayers. How can we explain the much stronger pressure dependency of the pull-off force (and thus of the apparent adhesion energy) seen in the high field case, shown in Figs. 3b and 5b, and summarized in Figs. 4 and 6, which is the most striking result of this study? Very recent work has shown that a transverse electric field of the same magnitude as in our experiments (in the high field case) induces electroporation and hydrophobic contacts between two PC LBs attached to gold and mica substrates [15-17]. Once these form due to such transmembrane *E*-fields, even at low applied pressures, they contribute to the adhesion as described above, and the stronger the pressure the more extensive the effect: this is because higher pressures lead to more dehydration of the LBs [44]. We thus attribute the strong adhesion hysteresis seen in the high field case in our system to this *E*-field-induced process initiating these topological changes. We emphasize that these structures form with little change in the bilayer thickness, which is indeed what we observe. At yet higher pressures we would expect hemifusion to occur for the near-liquid-phase DMPC-LBs, as earlier observed [45], with an abrupt expulsion of a bilayer; but this is not the case at the contact pressures of the present study, and indeed the lipid layers do not change their thickness significantly in our experiments.

We may now account also for the results showing adhesion hysteresis with increasing pressures also for the case of no electric fields, as observed in Figs. 3a and 5a and also in earlier studies [40-43]: the idea that hydrophobic contacts form between bilayers that are sufficiently compressed (and thus somewhat dehydrated relative to uncompressed LBs), can explain this relatively weak hysteresis. Once they form, they are likely to have an attractive contribution to the overall interaction as the surfaces separate on unloading, and this may account for the hysteretic behaviour and even the overall net adhesion as seen in Figs. 3a and 5a in the case of highest P_{max} where their formation is likely to be most marked. Such hydrophobic contact formation and resulting hysteresis may also be related to the sharp increase in sliding friction observed by Sorkin et al. earlier [9] between mica surfaces with attached DMPC-LBs, at loads similar to or higher than those corresponding to the highest red data points in Fig. 3b (but not at the lower loads corresponding to the blue data, where the sliding friction remained very low, indicating unperturbed DMPC-LBs, consistent with the absence of such contacts or hysteretic effects).

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5. Conclusions

The apparent adhesion energy (derived from the pull-off force via the contact mechanics relation) between surface-attached DMPC-lipid bilayers near their liquid phase, and between DSPC vesicles in their gel phase, are studied both with and without an externally-applied transverse electric field (*E*-field). We find that in the absence of an *E*-field the apparent interaction hysteresis on loading/unloading is pressure-independent at low contact pressures *P* but increases with *P* at higher pressures. However, when an *E*-field is applied, the apparent adhesion energy depends on the pressure already from low *P* values. This behaviour is attributed to fusion intermediates such as electro-pores and hydrophobic contacts: these latter may form at high enough pressures even with no *E*-field, but in the presence of an *E*-field they may form spontaneously even in the absence of pressure. Once formed they proliferate at higher pressures due to progressive dehydration of the LBs, thereby providing a *P*-dependent attractive component to the inter-bilayer interactions. Our results thus have a clear connection to membrane interactions in biological systems.

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Conflict of interests

The authors have no conflict of interests to declare.

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